

TECHNICAL NOTE

Rui M. Brito,¹ Ph.D.; Teresa Ribeiro,¹ B.Sc.; Rosa Espinheira,¹ B.Sc.; and Helena Geada,^{1,2} Ph.D.

South Portuguese Population Data on the Loci HLA-DQA1, LDLR, GYPA, HBGG, D7S8 and Gc

REFERENCE: Brito RM, Ribeiro T, Espinheira R, Geada H. South Portuguese population data on the loci HLA-DQA1, LDLR, GYPA, HBGG, D7S8 and Gc. *J Forensic Sci* 1998;43(5): 1031–1036.

ABSTRACT: Five South Portuguese Caucasian subpopulations were analyzed for the HLA-DQA1, LDLR, GYPA, HBGG, D7S8 and Gc loci. Genotype distributions for these loci did not deviate from Hardy-Weinberg expectations. The allele and genotype frequencies found have been compared with previously published data from North and Central Portugal. A total of 11 out of 138 chi-square comparisons of allele frequencies between different Portuguese populations showed a certain degree of divergence. Alentejo, Algarve, Madeira Island and Azores Islands populations might be considered as different groups in a database. For forensic casework, a composite South Portuguese Caucasian population database was obtained for estimating multiple locus profile frequencies using the six PCR-based loci studied.

KEYWORDS: forensic science, DNA typing, population genetics, South Portugal, polymerase chain reaction, HLA-DQA1, LDLR, GYPA, HBGG, D7S8, Gc

DNA profiling has rapidly become a routine technique in forensic laboratories. One of the PCR strategies that have been developed for forensic application is the reverse dot-blot analysis of HLA-DQA1 (1,2) and Polymarker (PM) systems (3). HLA-DQA1 is a well characterized PCR-based forensic system, located on chromosome 6. The PM system include five loci: on chromosome 19—Low Density Lipoprotein Receptor (LDLR), on chromosome 4—Glycophorin A (GYPA), on chromosome 11—Haemoglobin G Gammaglobin (HBGG), on chromosome 7—D7S8 and on chromosome 4—Group Specific Component (Gc).

For estimating multiple locus genetic profile in identity testing it is necessary to compile allele/genotype data from subpopulations that can allow a relevant estimate of any potential differences in subgroups appropriate to forensic casework analysis.

Substantial population data for HLA-DQA1 and PM loci exist in

¹ Forensic scientists, director and research supervisor, respectively, Forensic Biology Laboratory, Institute of Legal Medicine, Lisbon, Portugal.

² Assistant professor, Department of Legal Medicine, Faculty of Medicine, University of Lisbon, Lisbon, Portugal.

Received 7 Nov. 1997; and in revised form 2 Feb. 1998; accepted 3 Feb. 1998.

American (1–5), Asiatic (6–13), Australian (14,15) and European populations (16–21). There also have been some subpopulation data generated (22–31), in which different populations in relatively smaller geographical regions or ethnic groups were compared.

The South Portuguese Caucasian population can be divided into five regions: Estremadura, Alentejo, Algarve, Madeira Island and Azores Islands, based on specific culture and historical aspects. Figure 1 shows the geographic location of these regions. Some of these populations are relatively isolated and may offer some insight into genetic variation among subgroups.

This paper provides population genetic data on six PCR-based loci in South Portuguese Caucasian subpopulations and a composite South Portuguese Caucasian population and compares data with North and Central Portugal (28–31).

Materials and Methods

Population Samples

Whole blood samples were collected in EDTA tubes from a total of 514 healthy unrelated South Portuguese Caucasian individuals obtained from paternity investigation casework. Five different South Portugal regions (Fig. 1) were studied, namely Estremadura (including Ribatejo) (n = 187), Alentejo (n = 89), Algarve (n = 31), Madeira Island (n = 149) and Azores Islands (n = 58). All individuals and their parents were natives to their respective regions.

Blood samples were air-dried on cotton cloth and DNA was extracted by the Chelex method, previously described by Singer-Sam et al. (32).

Amplification and Typing

The DNA samples were amplified and typed for HLA-DQA1 by using the Amplitype HLA-DQA1 Forensic DNA Amplification and Typing Kit (Perkin Elmer Corporation, Norwalk, CT). For PM loci (LDLR, GYPA, HBGG, D7S8 and Gc), DNA samples were amplified and typed by using the Amplitype PM PCR Amplification and Typing Kit (Perkin Elmer Corporation, Norwalk, CT), according to the manufacturer's protocol. Amplification by PCR was carried out in a GeneAmp PCR System 2400 (Perkin-Elmer). One to 10 ng of DNA were used for PCR.

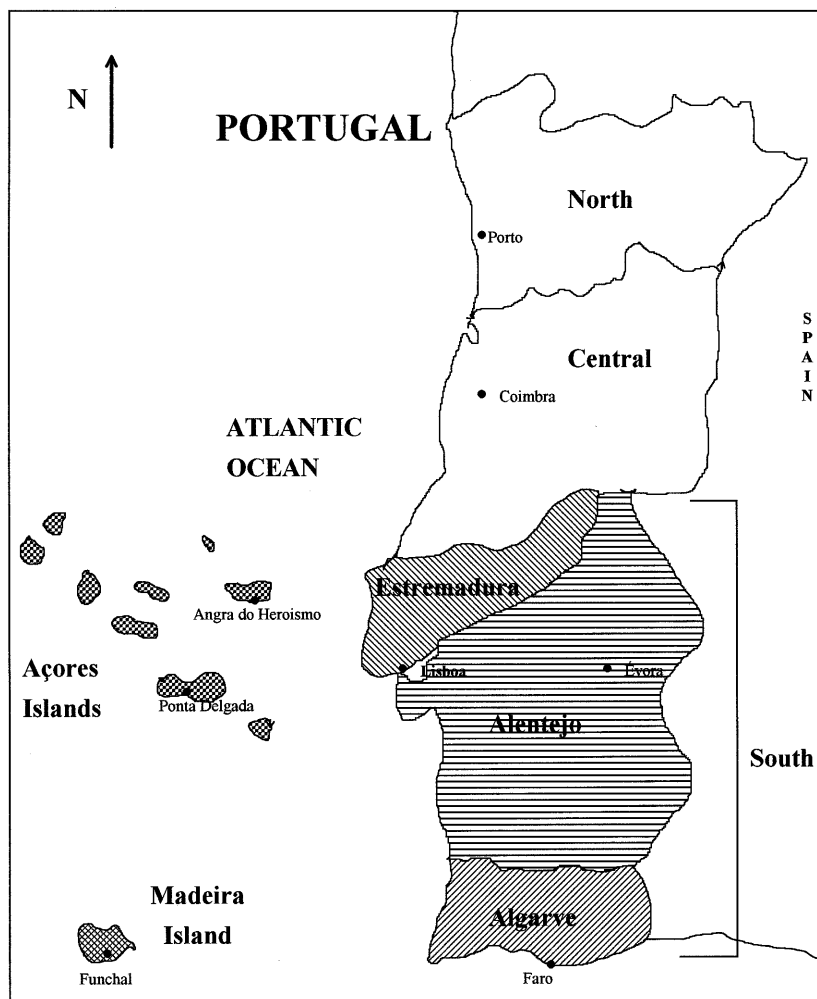


FIG. 1—Geographic location of the Portuguese regions: North, Central and South (Estremadura, Alentejo, Algarve, Madeira and Azores Islands).

Statistical Analysis

The distribution of the different genotype and allele frequencies for HLA-DQA1 and PM loci in the five different Portuguese subpopulation samples and for the total South Portugal sample was determined.

Departure from Hardy-Weinberg expectations was tested for each locus using conventional Pearson's chi-square method (χ^2). An exact significance probability test proposed by Sokal and Rohlf (33) was also performed.

Heterogeneity in allele frequencies for each locus was estimated for all pairwise comparisons of the populations groups by means of two-way RxC contingency table test and using a conventional χ^2 statistical parameter. Sampled populations were also compared with published data from North Portugal (28,29) and Central Portugal (30,31).

The potential usefulness of the studied markers for forensic studies in the Portuguese population was assessed by calculating the power of discrimination (PD), the chance of exclusion (CE) and the heterozygosity value (H).

The power of discrimination for each locus was calculated from genotype data, according to the formula given by Fisher (34). The chance of exclusion was calculated according to the formula given by Ohno et al. (35). The unbiased heterozygosity value was calculated using the formula described by Nei and Roychoudhury (36).

Results and Discussion

Genotype and Allele Frequencies

Genotype and allele frequencies for the HLA-DQA1 and PM loci in the five different Portuguese subpopulations and for the combined South Portugal population are shown in Tables 1–4.

The genotype frequencies for all loci did not deviate from Hardy-Weinberg expectations based on exact probability tests (Tables 1 and 3). By using the conventional χ^2 method, just three slight deviations from the Hardy-Weinberg equilibrium were observed due to an excess of heterozygotes at Gc in Estremadura and Alentejo samples and a deficit of heterozygotes at HLA-DQA1 in Alentejo. It should be noted that there would still be little impact for forensic purposes using these allele frequencies assuming independence, particularly because the exact probabilities did not show any deviations. In what concerns the combined South Portugal population, the genotype frequencies for the six loci did not deviate from Hardy-Weinberg expectations based on the conventional χ^2 method and on the exact probability tests.

The allele frequencies for the HLA-DQA1 and PM loci showed to be consistent with other European Caucasian populations.

Comparing the five Portuguese subpopulations, while Estremadura had the lowest value for HBGG allele C, Algarve had the highest values for the LDLR allele B, the HBGG allele C and the

Gc allele B. These data suggest that Algarve may have some African admixture as we will discuss later on.

Forensic Statistical Parameters

The forensic statistical parameters of the studied loci are presented in Table 5. In all five Portuguese subpopulations, HLA-DQA1 locus had a high heterozygosity ($H > 0.79$) and a high power of discrimination ($PD > 0.91$). For the PM system, the higher power of discrimination was obtained for the Gc locus, specifically, in the Algarve region ($PD = 0.786$), while the heterozygosity was just slightly higher ($H = 0.634$).

The combined power of discrimination for HLA-DQA1 and PM in the South Portugal population was 0.999; the combined chance of exclusion for the six loci studied in the same combined population was 0.885, while the mean heterozygosity value was 0.562, ranging from 0.477 for D7S8 to 0.809 for HLA-DQA1 (Table 6).

By the analysis of the statistical parameters obtained (PD , CE and H), it is shown that these markers are very useful for forensic application in all five subpopulations and in the combined South Portugal Caucasian population studied.

Comparison between Populations

Comparisons were performed between samples from the five Portuguese regions, and also with other Portuguese populations (28–31). A total of 11 out of 138 chi-square test performed were significant (Table 7). Differences were observed for LDLR, GYPA and HBG systems, mainly, when Alentejo and Algarve populations were compared with other Portuguese subpopulations. No significant differences were found for HLA-DQA1, D7S8 and Gc systems. When the total South Portugal population was compared with North and Central Portugal populations, no significant differences in allele frequencies were observed. These results suggest that a weighted construct of the South Portugal population to the actual population percentages would not be substantially different.

According to the results on HLA-DQA1 and PM loci, some Portuguese subpopulation data might be considered for estimating DNA profile frequencies. Alentejo, Algarve, Madeira and Azores Islands populations could be considered as independent groups in a database. However, the small number of differences observed had no statistical significance when the corrections for the multiple

TABLE 1—Observed genotype frequency distributions and Hardy-Weinberg equilibrium for HLA-DQA1 locus in the South Portuguese Caucasian populations.

Genotype	Estremadura (n = 182)	Alentejo (n = 81)	Algarve (n = 31)	Madeira Island (n = 149)	Azores Islands (n = 57)	South Portugal (n = 500)
1.1–1.1	0.011	0.025	0.032	0.027	0.052	0.024
1.1–1.2	0.033	0.049	0.032	0.040	0.052	0.040
1.1–1.3	0.028	0.062	0.032	0.040	0.018	0.036
1.1–2	0.055	0.012	0.032	0.060	0.000	0.042
1.1–3	0.039	0.062	0.065	0.027	0.070	0.044
1.1–4	0.099	0.074	0.161	0.121	0.105	0.106
1.2–1.2	0.017	0.025	0.032	0.034	0.018	0.024
1.2–1.3	0.033	0.037	0.000	0.040	0.018	0.032
1.2–2	0.039	0.000	0.032	0.060	0.035	0.038
1.2–3	0.028	0.025	0.065	0.000	0.035	0.022
1.2–4	0.071	0.161	0.194	0.067	0.123	0.098
1.3–1.3	0.006	0.049	0.032	0.013	0.018	0.018
1.3–2	0.039	0.012	0.000	0.013	0.035	0.024
1.3–3	0.033	0.000	0.000	0.013	0.018	0.018
1.3–4	0.065	0.049	0.000	0.047	0.052	0.052
2–2	0.043	0.025	0.000	0.047	0.035	0.038
2–3	0.043	0.062	0.032	0.040	0.035	0.044
2–4	0.126	0.111	0.065	0.114	0.105	0.114
3–3	0.022	0.025	0.000	0.034	0.018	0.024
3–4	0.077	0.049	0.129	0.094	0.123	0.086
4–4	0.093	0.086	0.065	0.067	0.035	0.076
χ^2	3.332	26.591	26.623	20.038	10.602	20.084
p	0.999	0.032	0.055	0.170	0.780	0.169
p^*	0.864	0.800	0.420	0.426	0.111	0.091

* Exact test probability.

TABLE 2—HLA-DQA1 observed allele frequencies in the South Portuguese Caucasian populations.

Allele	Estremadura (n = 182)	Alentejo (n = 81)	Algarve (n = 31)	Madeira Island (n = 149)	Azores Islands (n = 57)	South Portugal (n = 500)
1.1	0.137	0.154	0.194	0.171	0.175	0.158
1.2	0.118	0.160	0.194	0.138	0.149	0.139
1.3	0.104	0.130	0.048	0.091	0.088	0.099
2	0.195	0.123	0.081	0.191	0.140	0.169
3	0.132	0.123	0.145	0.121	0.158	0.131
4	0.313	0.309	0.339	0.289	0.289	0.304

TABLE 3—Observed genotype frequency distributions and Hardy-Weinberg equilibrium for PM loci in the South Portuguese Caucasian populations.

Locus	Genotype	Estremadura (n = 187)	Alentejo (n = 89)	Algarve (n = 31)	Madeira Island (n = 149)	Azores Islands (n = 58)	South Portugal (n = 514)
LDLR	AA	0.166	0.169	0.097	0.148	0.207	0.162
	AB	0.508	0.584	0.419	0.503	0.603	0.525
	BB	0.326	0.247	0.484	0.349	0.190	0.313
	χ^2	0.302	2.577	0.028	0.314	2.287	2.843
	<i>p</i>	0.582	0.108	0.867	0.575	0.130	0.092
	<i>p</i> *	0.653	0.136	1.000	0.611	0.188	0.104
GYPA	AA	0.262	0.360	0.193	0.295	0.224	0.280
	AB	0.508	0.528	0.581	0.443	0.448	0.490
	BB	0.230	0.112	0.226	0.262	0.328	0.230
	χ^2	0.039	1.268	0.662	2.021	0.609	0.165
	<i>p</i>	0.844	0.260	0.416	0.155	0.435	0.685
	<i>p</i> *	0.884	0.365	0.485	0.189	0.594	0.724
HBGG	AA	0.305	0.225	0.323	0.154	0.259	0.243
	AB	0.449	0.506	0.355	0.537	0.517	0.486
	AC	0.005	0.022	0.032	0.000	0.000	0.008
	BB	0.235	0.236	0.226	0.275	0.190	0.241
	BC	0.005	0.011	0.064	0.034	0.034	0.022
	CC	0.000	0.000	0.000	0.000	0.000	0.000
	χ^2	1.514	0.516	2.005	6.196	2.546	3.396
	<i>p</i>	0.679	0.915	0.571	0.102	0.467	0.335
	<i>p</i> *	0.241	0.834	0.279	0.068	1.000	0.791
	D7S8	AA	0.390	0.416	0.516	0.356	0.293
AB		0.428	0.438	0.323	0.503	0.500	0.453
BB		0.182	0.146	0.161	0.141	0.207	0.166
χ^2		2.189	0.327	2.425	0.404	0.001	1.286
<i>p</i>		0.139	0.568	0.119	0.525	0.993	0.257
<i>p</i> *		0.168	0.648	0.211	0.606	1.000	0.268
Gc	AA	0.086	0.045	0.000	0.054	0.035	0.058
	AB	0.043	0.135	0.161	0.107	0.103	0.091
	AC	0.337	0.427	0.387	0.309	0.310	0.344
	BB	0.011	0.000	0.032	0.040	0.034	0.021
	BC	0.219	0.123	0.226	0.181	0.172	0.188
	CC	0.304	0.270	0.194	0.309	0.345	0.298
	χ^2	8.069	8.746	4.356	1.390	1.249	5.012
	<i>p</i>	0.045	0.033	0.226	0.708	0.741	0.171
	<i>p</i> *	0.070	0.393	0.295	1.000	1.000	0.088

* Exact test probability.

TABLE 4—PM observed allele frequencies in the South Portuguese Caucasian populations.

Locus	Allele	Estremadura (n = 187)	Alentejo (n = 89)	Algarve (n = 31)	Madeira Island (n = 149)	Azores Islands (n = 58)	South Portugal (n = 514)
LDLR	A	0.420	0.461	0.306	0.399	0.509	0.424
	B	0.580	0.539	0.694	0.601	0.491	0.576
GYPA	A	0.516	0.624	0.484	0.517	0.448	0.525
	B	0.484	0.376	0.516	0.483	0.552	0.475
HBGG	A	0.532	0.489	0.516	0.423	0.517	0.490
	B	0.463	0.494	0.435	0.560	0.466	0.495
	C	0.005	0.017	0.048	0.017	0.017	0.015
D7S8	A	0.604	0.635	0.677	0.607	0.543	0.608
	B	0.396	0.365	0.323	0.393	0.457	0.392
Gc	A	0.275	0.326	0.274	0.262	0.241	0.276
	B	0.142	0.129	0.226	0.185	0.172	0.161
	C	0.583	0.545	0.500	0.554	0.586	0.563

typing were considered. Although there were differences in allele frequencies at some loci in some of the subgroups, an estimate of a six locus profile would be rare in all subgroups. More data are needed to determine whether or not the hypothesis that substantial differences in allele frequencies could affect forensic estimates.

One of the possible reasons for the significant differences

observed between Alentejo, Algarve and all the other subpopulations could be related to historical events. Alentejo and Algarve populations probably reflect the Arab occupation influence in Portugal until the 13th century, allied with a high degree of African migration during the 16th and 17th centuries, which might result in a potential African admixture. The observed differences between

TABLE 5—Statistical parameters of forensic value for HLA-DQA1 and PM loci in the South Portuguese Caucasian subpopulations.

Locus	Statistical Parameters	Estre-madura	Alentejo	Algarve	Madeira Island	Azores Islands
HLA-DQA1	PD	0.934	0.937	0.919	0.937	0.938
	CE	0.614	0.622	0.577	0.623	0.625
	H	0.805	0.813	0.793	0.812	0.818
LDLR	PD	0.618	0.622	0.577	0.614	0.623
	CE	0.184	0.187	0.167	0.182	0.185
	H	0.488	0.500	0.432	0.481	0.504
GYPA	PD	0.624	0.607	0.621	0.624	0.620
	CE	0.187	0.179	0.187	0.187	0.186
	H	0.501	0.472	0.508	0.501	0.499
HBGG	PD	0.631	0.648	0.684	0.644	0.647
	CE	0.193	0.249	0.243	0.206	0.208
	H	0.504	0.519	0.551	0.509	0.520
D7S8	PD	0.613	0.604	0.585	0.612	0.621
	CE	0.182	0.178	0.171	0.182	0.186
	H	0.480	0.466	0.444	0.479	0.501
Gc	PD	0.742	0.747	0.786	0.765	0.749
	CE	0.297	0.300	0.340	0.318	0.304
	H	0.566	0.583	0.634	0.593	0.573

PD = power of discrimination, CE = chance of exclusion, H = heterozygosity.

TABLE 6—Power of discrimination, change of exclusion and heterozygosity values for the six loci in the combined South Portugal Caucasian population.

Statistical Parameters	HLA-DQA1	LDLR	GYPA	HBGG	D7S8	Gc	Combined Values
PD	0.937	0.619	0.624	0.646	0.612	0.755	0.999
CE	0.622	0.185	0.187	0.206	0.182	0.309	0.885
H	0.809	0.489	0.499	0.515	0.477	0.581	0.562

PD = power of discrimination, CE = chance of exclusion, H = heterozygosity.

TABLE 7—Chi-square comparisons of allele frequencies between different Portuguese populations.

	HLA-DQA1	LDLR	GYPA	HBGG	D7S8	Gc
Estremadura/Alentejo	5.787	0.821	5.638**	2.458	0.475	1.493
Estremadura/Algarve	9.275	2.838	0.220	8.701**	1.201	3.088
Estremadura/Madeira	2.530	0.287	0.000	9.312**	0.007	2.258
Estremadura/Azores	3.776	2.835	1.626	1.573	1.370	0.945
Estremadura/Central Portugal	5.242	0.100	1.069	3.276	0.650	4.706
Estremadura/North Portugal	9.540	1.433	0.005	3.551	0.100	0.200
Alentejo/Algarve	4.496	4.487*	3.704*	2.231	0.365	3.350
Alentejo/Madeira	5.122	1.719	5.152*	1.990	0.356	3.701
Alentejo/Azores	2.090	0.647	8.738**	0.236	2.459	2.822
Alentejo/Central Portugal	2.779	0.321	1.762	0.040	1.704	0.848
Alentejo/North Portugal	7.118	0.032	4.980*	0.258	0.799	0.752
Algarve/Madeira	6.661	1.873	0.222	4.812	1.068	0.757
Algarve/Azores	2.954	6.708**	0.206	1.476	3.013	1.315
Algarve/Central Portugal	6.121	3.253	1.114	2.716	2.290	2.773
Algarve/North Portugal	6.885	5.287*	0.166	1.434	1.522	3.566
Madeira/Azores	2.167	4.069*	1.567	3.066	1.426	0.360
Madeira/Central Portugal	6.418	0.610	0.940	1.713	0.708	5.469
Madeira/North Portugal	3.945	2.613	0.007	3.909	0.140	2.857
Azores/Central Portugal	1.566	1.808	3.820	0.470	0.254	4.488
Azores/North Portugal	1.408	0.471	1.294	0.114	0.733	1.532
Central Portugal/North Portugal	7.918	0.639	0.986	0.520	0.190	3.298
South Portugal/Central Portugal	4.676	0.059	0.873	0.146	1.077	5.531
South Portugal/North Portugal	7.896	1.581	0.113	0.852	0.219	1.298

* Deviation at $p < 0.05$ level.

** Deviation at $p < 0.01$ level.

Madeira and the Azores Islands could be seen as a consequence of different colonizing populations together with different degrees of isolation.

For forensic purposes, a combined South Portuguese Caucasian population database including the five subpopulations studied can be generated for estimating multiple locus profile frequencies on HLA-DQA1 and PM loci.

Acknowledgments

The authors wish to express thanks to Dr. Bruce Budowle for comments on this manuscript prior to its submission.

The authors would like also to thank Isabel Lucas and Suzel Sousa for excellent technical assistance.

References

1. Comey CT, Budowle B. Validation studies on the analysis of the HLA DQ α locus using the polymerase chain reaction. *J Forensic Sci* 1991;36:1633–48.
2. Comey CT, Budowle B, Adams DE, Baumstark AL, Lindsey JA, Presley LA. PCR amplification and typing of the HLA DQ α gene in forensic samples. *J Forensic Sci* 1993;38(2):239–49.
3. Herrin G Jr, Fildes N, Reynolds R. Evaluation of the AmpliType® PM DNA test system on forensic case samples. *J Forensic Sci* 1994;39(5):1247–53.
4. Crouse CA, Feuer WJ, Nippes DC, Hutto SC, Barnes KS, Coffman D, et al. Analysis of HLA DQA1 allele and genotype frequencies in populations from Florida. *J Forensic Sci* 1994;39(3):731–42.
5. Budowle B, Lindsey JA, DeCou JA, Koons BW, Giusti AM, Comey CT. Validation and population studies of the loci LDLR, GYPA, HBGG, D7S8 and Gc (PM loci), and HLA-DQA1 using a multiplex amplification and typing procedure. *J Forensic Sci* 1995;40(1):45–54.
6. Koh CL, Benjamin DG. HLA-DQA1 genotype and allele frequencies in Malays, Chinese and Indians in the Malaysian population. *Hum Hered* 1994;44:150–5.
7. Al-Nassar KE, Mathew J, Thomas N, Fatania HR. HLA-DQA1 allele and genotype frequencies in a native Kuwaiti population. *Forensic Sci Int* 1995;72:65–9.
8. Hayes JM, Budowle B, Freund M. Arab population data on the

- PCR-based loci: HLA-DQA1, LDLR, GYPA, HBGG, D7S8, Gc and D1S80. *J Forensic Sci* 1995;40(5):888-92.
9. Huang NE, Budowle B. Chinese population data on the PCR-based loci HLA-DQ Alpha, Low-Density-Lipoprotein receptor, Glycophorin A, Haemoglobin G, D7S8, and Group-Specific Component. *Hum Hered* 1995;34-40.
 10. Tie J, Oshida S, Chiba S, Tsukamoto S, Sebetan IM. Frequency of D1S80 and HLA DQA1 alleles in a Chinese population. *Int J Legal Med* 1995;108:170-1.
 11. Woo KM, Budowle B. Korean data on the PCR-based loci LDLR, GYPA, HBGG, D7S8, Gc, HLA-DQA1 and D1S80. *J Forensic Sci* 1995;40(4):645-8.
 12. Alkhayat A, Alshamali F, Budowle B. Population data on the PCR-based loci LDLR, GYPA, HBGG, D7S8, Gc, HLA-DQA1 and D1S80 from Arabs from Dubai. *Forensic Sci Int* 1996;81:29-34.
 13. Nakajima T, Matsuki T, Ohkawara H, Nara M, Furukawa K, Kishi K. Evaluation of 7 DNA markers (HLA-DQA1, D1S80, LDLR, GYPA, HBGG, D7S8 and Gc) in a Japanese population. *Int J Legal Med* 1996;109:47-8.
 14. Gutowski S, Budowle B, Auer J, Oorschot RV. Statistical analysis of an Australian population for the loci Gc, HLA-DQA1, D1S80 and HUMTH01. *Forensic Sci Int* 1995;76:1-6.
 15. Stringer P, Triggs CM, Bakldwin LC, Melia MG, Savill MG. Distribution of HLA DQA1 alleles in a New Zealand Caucasian, Maori and Pacific Islander populations. Comparison with other population studies. *Int J Legal Med* 1995;108:2-7.
 16. Ambach E, Zehethofer K, Scheithauer R. HLA-DQA1 genotype and allele frequencies in an Austrian population. *Hum Hered* 1996;46:71-5.
 17. Cowland JB, Madsen HO, Morling N. HLA-DQA1 typing in Danes by two polymerase chain reactions (PCR) based methods. *Forensic Sci Int* 1995;73:1-13.
 18. Kloosterman AT, Sjerps M, Wust D. Dutch Caucasian population data on the loci LDLR, GYPA, HBGG, D7S8 and Gc. *Int J Legal Med* 1995;108:36-8.
 19. Budowle B, Woller J, Koons BW, Furedi S, Errera JD, Padar Z. Hungarian population data on seven PCR-based loci. *J Forensic Sci* 1996;41(4):667-70.
 20. Gehring C, Hochmeister MN, Budowle B, Reynolds R, Dirnhofer R. HLA-DQA1 subtyping data in the Swiss population. *Forensic Sci Int* 1996;83:27-30.
 21. Woller J, Budowle B, Furedi S, Padar Z. Hungarian population data on the loci HLA-DQA1, LDLR, GYPA, HBGG, D7S8 and Gc. *Int. J Legal Med* 1996;108:280-2.
 22. Allen M, Saldeen T, Pettersson U, Gyllensten U. Genetic typing of HLA class II genes in Swedish populations: applications to forensic analysis. *J Forensic Sci* 1993;38(3):554-70.
 23. Iriondo M, Manzano C, Rúa C. HLA-DQA1 in autochthonous Basques: description of a genocline for the DQA1*0201 allele in Europe. *Int J Legal Med* 1996;109:181-5.
 24. Keys KM, Budowle B, Andelinovic S, Definis-Gojanovic M, Drmic I, Mladen M, et al. Northern and Southern Croatian population data on seven PCR-based loci. *Forensic Sci Int* 1996;81:191-9.
 25. Martinez-Jarreta B, Bolea M, Castellano M, Hinojal R, Abecia E. Distribution of HLA DQA1 alleles and genotypes in two Spanish populations (Aragon and Asturias). *Forensic Sci Int* 1996;81:185-90.
 26. Walkinshaw M, Strickland L, Hamilton H, Denning K, Gayley T. DNA profiling in two Alaskan native populations using HLA-DQA1, PM and D1S80 loci. *J Forensic Sci* 1996;41(3):478-84.
 27. Scholl S, Budowle B, Radecki K, Salvo M. Navajo, Pueblo and Sioux population data on the HLA-DQA1, LDLR, GYPA, HBGG, D7S8, Gc and D1S80. *J Forensic Sci* 1996;41(1):47-51.
 28. Pinheiro MF, Pontes ML. The distribution of HLA DQA1 alleles in the population of the North of Portugal. In: Bar W, Fiori A, Rossi U, editors. *Advances on Forensic Haemogenetics*. Heidelberg: Springer-Verlag, 1993;559-61.
 29. Pinheiro MF, Pontes ML, Pinto-da-Costa J. Use of the amplitype PM coamplification system on forensic analysis. In: Margin P, Ledis B, editors. *Proceedings of XVIth Congress of the International Academy of Legal Medicine (Acta Medicinæ Legalis, Vol. XLIV)*, 1994;81-2.
 30. Rodríguez-Calvo MS, Bellas S, Souto L, Vide C, Valverde E, Carracedo A. Population data on the loci LDLR, GYPA, HBGG, D7S8, and Gc in three Southwest European populations. *J Forensic Sci* 1996;41(2):291-6.
 31. Lareu MV, Muñoz I, Pestoni C, Rodríguez MS, Vide C, Carracedo A. The distribution of HLA DQA1 and D1S80 (pMCT118) alleles and genotypes in the populations of Galicia and Central Portugal. *Int J Legal Med* 1993;106:124-8.
 32. Singer-Sam J, Tanguay RL, Riggs AD. Use of chelex to improve the PCR signal from a small number of cells. *Amplifications* 1989;3:11.
 33. Sokal RR, Rohlf FJ. *Biometry*, 2nd ed. W. H. Freeman and Company. San Francisco, 1981.
 34. Fisher RA. Standard calculations for evaluating a blood group system. *Heredity* 1951;5:95-102.
 35. Ohno Y, Sebetan IM, Akaishi S. A simple method for calculating the probability of excluding paternity with any number of codominant alleles. *Forensic Sci Int* 1982;19:93-8.
 36. Nei M, Roychoudhury AK. Sampling variances of heterozygosity and genetic distance. *Genetics* 1974;76:379-90.

Additional information and reprint requests:
 Helena Geada, Ph.D.
 Institute of Legal Medicine
 Rua Manuel Bento de Sousa, 3
 1150 Lisbon
 Portugal